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## Erythromycin Delayed-Release Tablets

### DEFINITION

Erythromycin Delayed-Release Tablets contain NLT 90.0% and NMT 120.0% of the labeled amount of erythromycin ( $C_{37}H_{67}NO_{13}$ ).

### IDENTIFICATION

- **A. THIN-LAYER CHROMATOGRAPHY**

**Standard solution:** 2.5 mg/mL of [USP Erythromycin RS](#) in [methanol](#)

**Sample solution:** Nominally 2.5 mg/mL of erythromycin from powdered Tablets in [methanol](#)

**Chromatographic system**

(See [Chromatography \(621\), General Procedures, Thin-Layer Chromatography](#).)

**Mode:** TLC

**Adsorbent:** 0.25-mm layer of chromatographic silica gel

**Application volume:** 10  $\mu$ L

**Developing solvent system:** [Methanol](#) and [chloroform](#) (85:15)

**Spray reagent:** [Alcohol](#), *p*-methoxybenzaldehyde, and [sulfuric acid](#) (90:5:5)

**Analysis**

**Samples:** Standard solution and Sample solution

Place the plate in an unlined chromatographic chamber, and develop the chromatogram until the solvent front has moved about 7 cm.

Remove the plate from the chamber, mark the solvent front, and allow the solvent to evaporate. Spray the plate with Spray reagent. Heat the plate at 100° for 10 min, and examine the chromatogram, in which erythromycin appears as a black-to-purple spot.

**Acceptance criteria:** The  $R_F$  value of the principal spot of the Sample solution corresponds to that of the Standard solution.

### ASSAY

- [ANTIBIOTICS—MICROBIAL ASSAYS \(81\)](#)

**Sample solution:** Place NLT 4 Tablets in a high-speed glass blender jar with 200 mL of [methanol](#), and blend for 3 min. Add 300 mL of *Buffer B.3*, and blend for 3 min.

**Analysis:** Proceed as directed in the chapter. Dilute the Sample solution with *Buffer B.3* to obtain a *Test Dilution* having a concentration that is nominally equivalent to the median level of the standard.

**Acceptance criteria:** 90.0%–120.0%

### PERFORMANCE TESTS

**Change to read:**

- [DISSOLUTION \(711\)](#)

**Test 1:** If the product complies with this test, the labeling indicates that it meets USP Dissolution Test 1.

Proceed as directed in [Dissolution \(711\), Procedure, Apparatus 1 and Apparatus 2, Delayed-Release Dosage Forms, Method B Procedure](#).

**Acid stage**

**Medium:** [Simulated gastric fluid TS](#), without pepsin; 900 mL

**Apparatus 1:** 100 rpm

**Time:** 60 min

**Analysis:** Do not analyze the sample at this stage.

**Buffer stage**

**Medium:** 0.05 M pH 6.8 phosphate buffer (see [Reagents, Indicators, and Solutions—Buffer Solutions](#))<sup>▲</sup>; 900 mL<sup>▲</sup> (ERR 1-Apr-2023)

**Apparatus 1:** 100 rpm

**Time:** 60 min

**Buffer:** pH 1.2 buffer (see [Reagents, Indicators, and Solutions—Buffer Solutions](#))

**Solution A:** 1 g/L of [bromocresol purple](#) in pH 4.5 phosphate buffer

**Standard solution:** Dissolve [USP Erythromycin RS](#) in Medium to obtain a concentration similar to that of the Sample solution.

**Sample solution:** If necessary, dilute a filtered portion of the solution under test with Medium to obtain a solution containing about 0.28 mg/mL of erythromycin.

**Detector:** UV 410 nm

**Analysis**

**Samples:** Standard solution and Sample solution

Transfer 2.0 mL of the *Standard solution* and the *Sample solution* to individual separators of a suitable size. Add 6 mL of *Buffer* and 8 mL of *Solution A*, and mix. Extract with 40.0 mL of chloroform. Determine the amount of erythromycin ( $C_{37}H_{67}NO_{13}$ ) dissolved from UV absorbances of the chloroform extracts.

**Tolerances:** NLT 75% (Q) of the labeled amount of erythromycin ( $C_{37}H_{67}NO_{13}$ ) is dissolved.

**Test 2:** If the product complies with this test, the labeling indicates that it meets USP *Dissolution Test 2*. Proceed as directed under *Test 1*, except to use *Apparatus 2* at 75 rpm.

**Test 3:** If the product complies with this test, the labeling indicates that it meets USP *Dissolution Test 3*.

**Acid stage**

**Acid stage medium:** [Simulated gastric fluid TS](#), without enzyme; 900 mL

**Apparatus 1:** 100 rpm

**Time:** 60 min

**Solution A:** 3.6 g/L of [dibasic sodium phosphate](#) in [water](#). Adjust with [diluted phosphoric acid](#) to a pH of 9.0.

**Mobile phase:** *Solution A* and [acetonitrile](#) (1:1)

**Solution B:** 6.8 g/L of [monobasic potassium phosphate](#) and 1.2 g/L of [sodium hydroxide](#) in [water](#)

**Peak identification solution:** 0.05 mg/mL of [USP Erythromycin B RS](#) and [USP Erythromycin C RS](#) prepared as follows. Transfer 2.5 mg each of [USP Erythromycin B RS](#) and [USP Erythromycin C RS](#) to a 50-mL volumetric flask, add 12.5 mL of [methanol](#), sonicate to dissolve, and dilute with *Solution B* to volume.

[*NOTE*—The typical retention times of erythromycin C and erythromycin B are 4.2 and 13.4 min, respectively.]

**Standard solution:** 2.5 mg/mL of [USP Erythromycin RS](#) prepared as follows. Transfer 125 mg of [USP Erythromycin RS](#) to a 50-mL volumetric flask, add 12.5 mL of [methanol](#), sonicate to dissolve, and dilute with *Solution B* to volume.

[*NOTE*—The typical retention time of erythromycin A is 7.8 min.]

**Sample solution 1:** Determine the average Tablet weight by weighing NLT 20 Tablets. Carefully transfer the appropriate number of intact Tablets into a suitable volumetric flask (5 Tablets into a 500-mL flask for 250-mg Tablets, 8 Tablets into a 1000-mL flask for 333-mg Tablets, and 5 Tablets into a 1000-mL flask for 500-mg Tablets). Add [methanol](#) to about 25% of the final volume, and sonicate at room temperature for about 30 min with intermittent shaking. Further add about 25% of the final volume of *Solution B* and sonicate at room temperature for about 30 min with intermittent shaking. Dilute to volume with *Solution B* and mix well. Centrifuge at 5000 rpm for 5 min and pass the supernatant through a polyvinylidene fluoride (PVDF) or other suitable filter of 0.45- $\mu$ m pore size. Discard the first 5 mL of the filtrate.

**Sample solution 2:** At the end of *Acid stage* dissolution, discard *Acid stage medium* and carefully transfer 1 Tablet from the dissolution vessel into a suitable volumetric flask (use a 100-mL flask for 250-mg Tablets, 200-mL flask for 333-mg Tablets, and 200-mL flask for 500-mg Tablets). Add [methanol](#) to about 25% of the final volume, and sonicate at room temperature for about 30 min with intermittent shaking. Further add about 25% of the final volume of *Solution B* and sonicate at room temperature for about 30 min with intermittent shaking. Dilute to volume with *Solution B* and mix well. Centrifuge at 5000 rpm for 5 min and pass the supernatant through a PVDF or other suitable filter of 0.45- $\mu$ m pore size. Discard the first 5 mL of the filtrate.

**Blank:** *Solution B* and methanol (75:25)

**Chromatographic system**

(See [Chromatography \(621\), System Suitability](#)).

**Mode:** LC

**Detector:** UV 215 nm

**Column:** 4.6-mm x 25-cm; 5- $\mu$ m packing L1

**Temperature**

**Autosampler:** 4°

**Column:** 50°

**Flow rate:** 1.5 mL/min

**Injection volume:** 25  $\mu$ L

**System suitability**

**Sample:** Standard solution

[*NOTE*—The relative retention times of erythromycin C, erythromycin A, and erythromycin B are 0.53, 1.00, and 1.75, respectively.]

**Suitability requirements**

**Tailing factor:** NMT 2.0 for erythromycin A peak

**Relative standard deviation:** NMT 2.0% of the sum of erythromycin A, erythromycin B, and erythromycin C

**Analysis**

**Samples:** Standard solution, Sample solution 1, and Sample solution 2

Calculate the erythromycin content (A) as a percentage of the labeled amount of erythromycin:

$$\text{Result} = (r_u/r_s) \times W \times P \times (1/D_s) \times D_1 \times (1/L) \times 100$$

$r_u$  = peak response of sum of erythromycin A, erythromycin B, and erythromycin C from Sample solution 1

$r_s$  = peak response of sum of erythromycin A, erythromycin B, and erythromycin C from the Standard solution

*W* = standard weight of [USP Erythromycin RS](#) to prepare the *Standard solution* (mg)

*P* = content of erythromycin A, erythromycin B, and erythromycin C in [USP Erythromycin RS](#) (mg/mg)

*D<sub>S</sub>* = dilution factor used in preparing the *Standard solution* (mL)

*D<sub>1</sub>* = dilution factor used in preparing *Sample solution 1* (mL)

*L* = label claim (mg/Tablet)

Calculate the percentage (*T*) of the labeled amount of erythromycin retained:

$$\text{Result} = (r_U/r_S) \times W \times P \times (1/D_S) \times (1/L) \times D_2 \times 100$$

*r<sub>U</sub>* = peak response of sum of erythromycin A, erythromycin B, and erythromycin C from *Sample solution 2*

*r<sub>S</sub>* = peak response of sum of erythromycin A, erythromycin B, and erythromycin C from the *Standard solution*

*W* = standard weight of [USP Erythromycin RS](#) to prepare the *Standard solution* (mg)

*P* = content of erythromycin A, erythromycin B, and erythromycin C in [USP Erythromycin RS](#) (mg/mg)

*D<sub>S</sub>* = dilution factor used in preparing the *Standard solution* (mL)

*L* = label claim (mg/Tablet)

*D<sub>2</sub>* = dilution factor used in preparing *Sample solution 2* (mL)

Calculate the percentage of the labeled amount of erythromycin dissolved in *Acid stage*:

$$\text{Result} = A - T$$

*A* = erythromycin content as a percentage of the labeled amount

*T* = percentage of the labeled amount of erythromycin retained

[**NOTE**—If *T* is greater than *A*, consider the result to be zero.]

**Tolerances:** NMT 10% of the labeled amount of erythromycin is dissolved.

#### Buffer stage

**Buffer stage medium:** 6.8 g/L [monobasic potassium phosphate](#) in water with pH 6.8 adjusted by 5 N [sodium hydroxide](#); 900 mL

**Apparatus 1:** 100 rpm

**Time:** 35 min

**Solution A and Mobile phase:** Prepare as directed in *Acid stage*.

**Standard solution:** Transfer a suitable amount of [USP Erythromycin RS](#) into an appropriate volumetric flask. See [Table 1](#). Add [methanol](#) to about 5% of the final volume and sonicate to dissolve. Dilute with *Buffer stage medium* to volume with intermittent shaking and mix well. [**NOTE**—The typical retention time of erythromycin A is 3.8 min.]

**Table 1**

Tablet Label Claim (mg)	Weight of <a href="#">USP Erythromycin RS</a> (mg)	Volumetric Flask (mL)
250	59	200
333	39	100
500	59	100

**Sample solution:** Prepare as directed in *Acid stage* with a new set of Tablets. After 60 min with *Acid stage medium*, immediately replace with *Buffer stage medium*. After 35 min, pass a portion of the solution through a PVDF or other suitable filter of 0.45-μm pore size.

#### Chromatographic system

(See [Chromatography \(621\), System Suitability](#).)

**Mode:** LC

**Detector:** UV 210 nm

**Column:** 4.6-mm x 15-cm; 5-μm packing L1

**Temperature**

**Autosampler:** 5°

**Column:** 50°

**Flow rate:** 2.0 mL/min**Injection volume:** 100 µL**System suitability****Sample:** Standard solution**Suitability requirements****Tailing factor:** NMT 2.0 for erythromycin A peak**Relative standard deviation:** NMT 2.0% of erythromycin A**Analysis****Samples:** Standard solution and Sample solution

Calculate the percentage of the labeled amount of erythromycin dissolved:

$$\text{Result} = (r_U/r_S) \times C_S \times (1/L) \times V \times 100$$

 $r_U$  = peak response of erythromycin A from the Sample solution $r_S$  = peak response of erythromycin A from the Standard solution $C_S$  = concentration of erythromycin A in the Standard solution (mg/mL) $L$  = label claim (mg/Tablet) $V$  = volume of buffer medium**Tolerances:** NLT 80% (Q) of the labeled amount of erythromycin is dissolved.

- **Uniformity of Dosage Units (905):** Meet the requirements

**SPECIFIC TESTS**

- **WATER DETERMINATION (921), Method I**

**Analysis:** Use 20 mL of methanol containing 10% of imidazole in place of methanol in the titration vessel.**Acceptance criteria:** NMT 6.0%**ADDITIONAL REQUIREMENTS**

- **PACKAGING AND STORAGE:** Preserve in tight containers.
- **LABELING:** The labeling indicates the *Dissolution Test* with which the product complies.
- **USP REFERENCE STANDARDS (11):**

USP Erythromycin RSUSP Erythromycin B RSUSP Erythromycin C RS**Auxiliary Information** - Please [check for your question in the FAQs](#) before contacting USP.We apologize for the inconvenience. The exact auxiliary information for this Documentary Standard is currently unavailable. Please contact Documentary Standards Support ([stdsmonographs@usp.org](mailto:stdsmonographs@usp.org)) for assistance during this time.**Chromatographic Database Information:** [Chromatographic Database](#)**Most Recently Appeared In:**

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